



The impact of electromagnetic radio waves on some biological aspects of *Culex (Culex) pipiens* Mosquitoes (Diptera: Culicidae)

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Experiments were conducted to investigate the impact of Radio-Frequency (RF) exposure on some biological aspects of immature stages of *Culex (Culex) pipiens* mosquito. Immature stages of *Cx. (Cx.) pipiens* mosquito were collected from Giza, Egypt and maintained at the Dept. of Entomology, Fac. of Science, Cairo University. 100 first instar larvae were exposed to a single dose of discontinuous RF using GSM multiband mobile phone for 4 hours simulating phone conversation conditions. Immature duration, percentage mortality, adult emergence, and sex ratio were calculated and statistically analyzed. RF-exposed insects exhibited significant reduction in developmental duration (19 and 10.55 days for control and RE-exposed, respectively), significant increase in percentage mortality (23.8, 11.2% and 38.6, 48.8% for control and RE-exposed larval and pupal mortality, respectively) and male-biased sex ratio (1♀: 1.34♂ and 1♀: 3.46♂ for control and RE-exposed, respectively). These results provided an evidence for the negative impact of RF-radiation on the components of ecosystem. Additional research on the molecular effects of RF-exposure on living organisms is encouraged.

Keywords: Radio-Frequency exposure, *Culex (Culex) pipiens*, mosquito, Diptera, Insects

INTRODUCTION

Human beings are generally uncovered from the effect of both mechanical and electromagnetic waves. Unlike mechanical waves (MW), electromagnetic (EM) waves propagate in vacuum. Electromagnetic spectrum includes a wide range of wavelengths from radio waves to gamma-rays (Reed, 2018, Balmori, 2021). Mobile phone cell is an artificial EM source (WHO, 2002, Perrin and Souques, 2012, Wargo et al. 2012). A considerable increase in the number of mobile phone devices was determined worldwide. In 2021, more than 8.6 billion subscriptions of mobile phone were reported. It was less than one billion subscriptions in 2000 (ITU, 2021). Consequently, the infrastructures like telecom companies' towers and telecom relay stations were installed. Several infrastructures were installed closer to densely populated urban residential areas (e.g. schools, nurseries, public playgrounds, commercial buildings, hospitals, university campuses, and amphitheatres) (Feynman, 2013). The increasing number of smart phones' users, in addition to the revolution of smart mobile technology that supports games, has led to the emergence of many studies confirming the impact of mobile phone use on the behavior of children and adolescents (e.g. Hardell, 2017, Kopecký et al. 2021). Thus, many risk-assessment studies

investigated the effect of mobile phone radiation on living organisms (Adey, 1975a, Bawin et al. 1975, Dutta et al. 1984, Adey, 1988, Goodman et al. 1995, Velizarov et al. 1999, Xenos and Margas, 2003, Cucurachi et al. 2013).

As a considerable component of biomass and ecosystem, insects are very important for the ecosystem sustainability and human life continuation (Samways, 1993, Miller and Spoolman, 2012). The impact of electromagnetic field (EMF) on insect was investigated by many researchers: development dynamics of *Musca domestica* (Stanojević et al. 2005); reproductive capacity of *Drosophila melanogaster* (Panagopoulos et al. 2004, 2007); ant sites cues (Cammaertset al. 2012); the generation time of *D. melanogaster* N strain (Fauzi et al. 2015); the generation time, sex ratio and filial number of *D. melanogaster* bony strain (Fauzi and Corebima, 2015); the development and performance of *Callosobruchuschinensis* (Coleoptera) (Maharjan et al. 2019a), oviposition preference and development of *Maruca vitrata* (Lepidoptera) (Maharjan et al. 2019b), life-table parameters of perilla seed bugs (*Nysius* sp.) (Heteroptera: Lygaeidae) (Maharjan et al. 2020); and survival rate and reproductive organs morphology of *D. melanogaster* (Sudaryadi et al. 2020). Such potential effects may result in dramatic decline of insect populations

(Balmori, 2009, 2014, 2015, 2021, Hallmann, 2017, Thill, 2020). Cellular phone radiations were also incriminated in the changes of behavior and colony collapse disorder of honey bee in USA (Kimmel et al. 2007, Mixsonet al. 2009, Sharma and Kumar, 2010, Kumar et al. 2011, Favre, 2011, Sainudeen, 2011, Mal and Kumar, 2014, Taye et al. 2017, Kumar, 2018, Favre and Johansson, 2020, Di Noi et al. 2021).

The present study aimed to investigate the impact of Radio Frequency (RF) radiation emitted by cellular mobile phone on larval *Culex (Cx.) pipiens* mosquito..

MATERIALS AND METHODS

Mosquito rearing

Larvae of *Cx. (Cx.) pipiens* were originally collected on October to September, 2020 from a breeding site in Giza, Egypt. This colony was maintained in the insectary of the Department of Entomology, Faculty of Science, Cairo University under controlled conditions (12Light: 12Dark photoperiod, 27 ± 1 °C temperature, and 70-80% RH). Larvae were fed with activated yeast. Adult mosquitoes were fed on the blood of tied domestic pigeon (*Columba liviademestica*) placed on the top side of the mosquito cages twice a week as previously described (Galalet al. 2017).

Radio-Frequency irradiation and experimental setup

One hundred newly hatched first instar larvae of *Cx. (Cx.) pipiens* were exposed to a single dose of discontinuous radio frequency (RF) signal produced by a 4G GSM multiband mobile phone (about 900/ 1900 MHz and power approximately 0.03 m W/cm²) for 4 hours under controlled conditions. To simulate a phone conversation, one mobile device called and the second answered. The two devices were in a continuous conversation during the RF-exposure period. Each device was hung up in front of and touching a glass jar (10-cm diameter x 20-cm height). Each jar contains 50 newly hatched first instar larvae. Control group was subjected to all conditions, but free from RF-exposure. Following the end point of the irradiation, the 100 larvae were divided into 10 cups (10 larvae/ cup) to facilitate observation recordings. Immature duration, percentage mortality, adult emergence, and sex ratio were observed and calculated. Four readings per day were taken during the first 24 hours post-irradiation, followed by one reading per day until the end of the experimental period. All experiments were repeated three times.

Statistical analyses

Statistical analyses were done using SPSS Ver. 25.0 software (SPSS Inc., Chicago, IL). Comparisons between means were done by using *t-test* at the significance level $P < 0.05$. Additionally, correlation analyses of duration-stage and mortality-stage were done for both control and RF-

exposed insects. Sex ratio data were subjected to *Chi²* test.

RESULTS

The effect of RF-exposure on the developmental period of immature stages of *Cx. (Cx.) pipiens*

The effect of RF-exposure on the developmental period of immature stages of *Cx. (Cx.) pipiens* was illustrated in Figure (1). Durations of the RF-exposed larvae were reduced to ≈half the value when compared to controls (RF-unexposed). Statistical analyses revealed that durations of the RF-exposed second larval, third larval, fourth larval, total larval, and total immature (larval+pupal) were significantly reduced ($P < 0.05$) when compared to control. Duration differences between RF-exposed and controls were insignificant ($P > 0.05$) in the cases of the first larval and pupal durations (Figure 1).

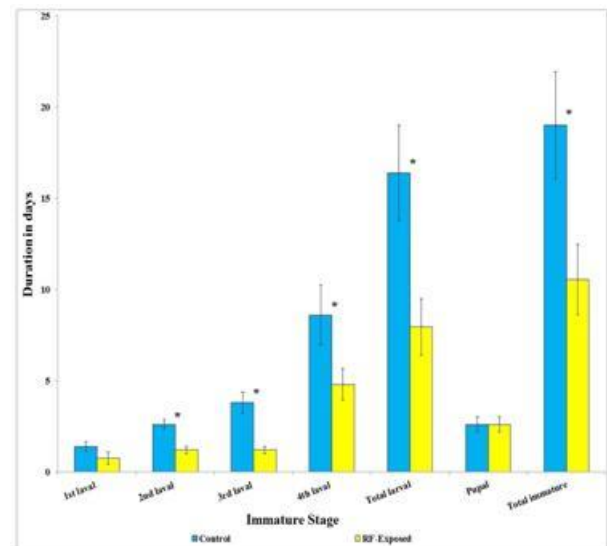


Figure 1: Effect of RF-exposure on the immature duration of *Cx. (Cx.) pipiens*. Asterisks refer to significant difference between RF-exposed insects and control at $P < 0.05$ using *t-test*.

Correlation analysis indicated that there were positive correlations between duration and immature stage in both RE-exposed and control insects (Figure 2). The correlation was weak positive ($R^2 = 0.49$) in control, and strong positive ($R^2 = 0.62$) in RF-exposed insects (Figure 2).

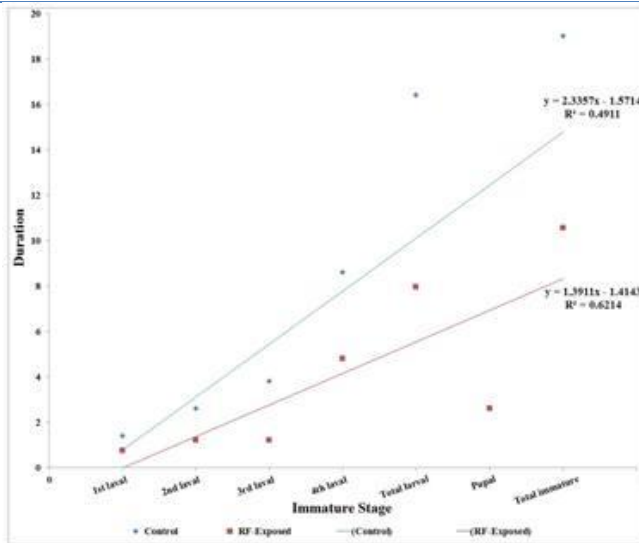


Figure 2. Duration-stage correlation of RF-exposed and control *Cx. (Cx.) pipiens*. $R^2 = 0.49$ for control (weak correlation), and $R^2 = 0.62$ for RF-exposed (strong correlation).

The effect of RF-irradiation on the percentage mortality of immature stages of *Cx. (Cx.) pipiens*

The effect of RF-exposure on the percentage mortality of immature stages of *Cx. (Cx.) pipiens* were illustrated in Figure (3). Percentage mortality of the RF-exposed larvae were increased to ≈double the value when compared to controls. Statistical analyses revealed that mortalities of the RF-exposed first larval, second larval, fourth larval, total larval, and pupal stages were significantly increased ($P < 0.05$) when compared to control mortality. Mortality difference between RF-exposed and control was insignificant ($P > 0.05$) in the case of the third larval stage (Figure 3).

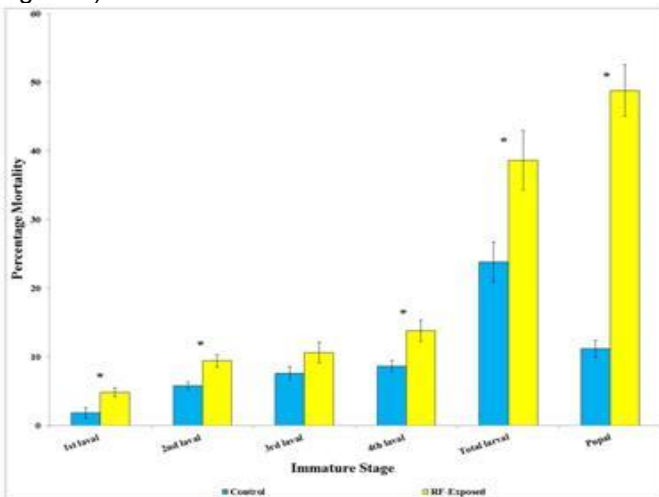


Figure 3: Effect of mobile radiation on the immature mortality of *Cx. (Cx.) pipiens*. Asterisks refer to significant difference between RF-exposed insects and control at $P < 0.05$ using *t*-test.

Correlation analysis indicated that there were positive

correlations between percentage mortality and immature stage in both RF-exposed and control insects (Figure 4). The correlation was strong positive ($R^2 = 0.84$) in control, and moderate positive ($R^2 = 0.53$) in RF-exposed insects (Figure 4). Additionally, RF-exposure led to increased number of adults' failure to emerge from pupae.

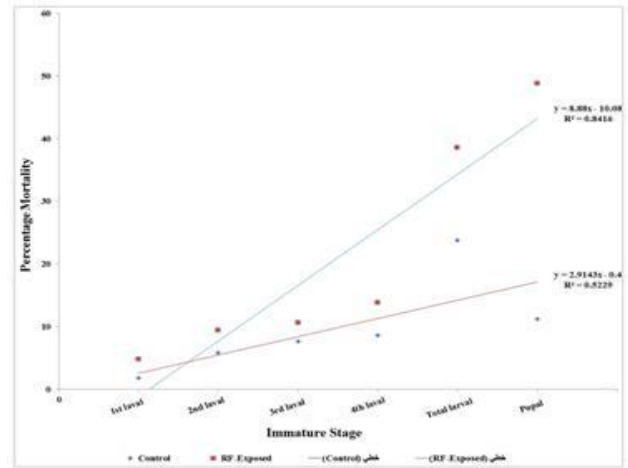


Figure 4: Mortality-stage correlation of RF-exposed and control *Cx. (Cx.) pipiens*. $R^2 = 0.84$ for control (strong correlation), and $R^2 = 0.52$ for RF-exposed (moderate correlation).

The effect of RF-irradiation on the sex ratio of *Cx. (Cx.) pipiens*

The effect of RF-exposure on the sex ratio of *Cx. (Cx.) pipiens* were illustrated in Figure (5).

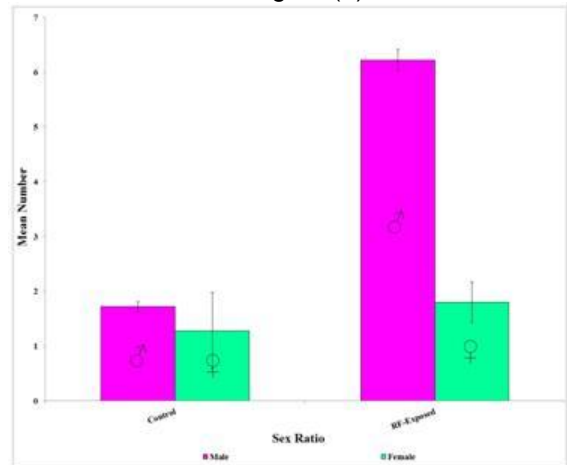


Figure 5: Effect of RF-exposure on the sex ratio of RF-exposed and control *Cx. (Cx.) pipiens*.

Sex ratio of the RF-exposed insects was male-biased ($1♀ : 3.46♂$) when compared to controls ($1♀ : 1.34♂$). Increase in the number of males emerged from RF-exposed insects was statistically significant ($P = 0.00$) when compared to controls (*t*-test). However the difference in the number of females was insignificant ($P = 0.25$) when compared to controls (*t*-test). χ^2 statistical

analysis of the sex ratio revealed that the difference in sex ratios of both RF-exposed and control insects was insignificant ($X^2= 18.0$ and $P= 0.08$).

DISCUSSION

The huge number of mobile phone devices, telecom towers, and relay stations emit a lot of EMF radiations within the ecosystem. The present study aimed to investigate the effect of RF irradiation on the immature stages of *Cx. (Cx.) pipiens*. Significant reduction was recorded in the developmental time of RE-exposed immature insects when compared to controls. This reduction was strongly correlated to RF-exposure. Contrarily, 50 Hz EMF-exposed *M. domestica* pupae showed significant retardation of metamorphosis (Stanojević et al. 2005). Additionally, significant elongated generation time of the first generation of two different strains of *D. melanogaster* was reported due to EMF-exposure when compared to control (Fauzi and Corebima, 2015, Fauzi et al. 2015). Furthermore, significant elongated developmental time was demonstrated for RF-treated *M. vitrata* when compared to control (Maharjan et al. 2019b). It is well-known that the shorter generation duration, the greater the insect population (Schowalte, 2006). The contradiction of the above-mentioned results and our results might correspond to the dose of irradiation, the irradiated stage and/ or the insect species. Furthermore, the effect of EMF on the generation time might correlate to the cellular and/ or hormonal processes of gonad development (Panagopoulos et al. 2004), or due to the concentration of ecdysteroid hormone (Gilbert et al. 2002, Atli and Unlu, 2007).

Another factor-influencing population is the percentage mortality. Significant increase was recorded in the percentage mortality of RE-exposed immature insects when compared to controls. This increase was moderately correlated to RF-exposure. As the effects of RF-exposure are non-thermal, the percentage mortality of RE-exposed insects might correlate to some proteins induced by the RF-exposure (Seufi et al. 2007, Wargo et al. 2012). These proteins might affect cellular protein functions (Weisbord et al. 2003). Consequently, RF-exposure could increase the failure of adults to emerge from pupae.

The last tested variable was sex ratio. We found that sex ratio of RF-exposed insects was significantly male-biased. Whereas, the Ch^2 test clarified that the difference in sex ratios of both control and RF-exposed insects was insignificant. Similarly, Mirabolghasemi and Azarnnia (2002) and Fauzi and Corebima (2015) reported insignificant effect of EMF exposure on sex ratio of the first generation *D. melanogaster* when compared to control. Generally, sex ratio of most species tends to be 1♀: 1♂ (Fisher's principle), however in some cases, sex ratio bias in favor of male or female may happen in favor of the entire population fitness (Hamilton, 1967, Edwards, 1998).

CONCLUSION

The present study demonstrated that the RF-exposed insects exhibited reduced developmental duration, increased percentage mortality and male-biased sex ratio. Therefore, our findings provided additional evidence to the adverse impacts of RF-radiation on the components of ecosystem. Additional research on the molecular effects of RF-exposure on living organisms is encouraged.

CONFLICT OF INTEREST

The authors declared that present study was performed in absence of any conflict of interest.

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AUTHOR CONTRIBUTIONS

GFH designed and performed practical work, data analysis and also wrote the manuscript. SAM participate in practical work, data analysis, writing the MS draft and reviewed the final manuscript. All authors read and approved the final version.

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REFERENCES

- Adey WR. 1988. Electromagnetic Field Interactions in the Brain. In: Başar E. (eds) Dynamics of Sensory and Cognitive Processing by the Brain. Springer Series in Brain Dynamics, vol 1. Springer, Berlin, Heidelberg. https://doi.org/10.1007/978-3-642-71531-0_9.
- Adey WR. 1975. Introduction: Effects of electromagnetic radiation on the nervous system. Ann. NY Acad. Sci. 247:1520- 1543.
- Atli E, Unlu H. 2007. The Effects of Microwave Frequency Electromagnetic Fields on the Fecundity of *Drosophila melanogaster*. Turk. J. Biol. 31:1- 5.
- Balmori A. 2021. Electromagnetic radiation as an emerging driver factor for the decline of insects. Sci. Total Environ. 767: 144913. doi:10.1016/j.scitotenv.2020.144913.
- Balmori A. 2015. Anthropogenic radiofrequency electromagnetic fields as an emerging threat to wildlife orientation. Sci. Total Environ. 518:58- 60.
- Balmori A. 2014. Electrosmog and species conservation. Sci. Total Environ. 496: 314-316.
- Balmori A. 2009. Electromagnetic pollution from phone

- masts. Effects on wildlife. *Pathophysiol.* 16(2-3): 191-199.
- Bawin SM, Kaczmarek LK, Adey WR. 1975. Effects of modulated VMF fields on the central nervous system. *Ann. NY Acad. Sci.* 247:74- 81. <http://dx.doi.org/10.1111/j.1749-6632.1975.tb35984.x>.
- Cammaerts MC, De Doncker P, Patris X, Bellens F, Rachidi Z, Cammaerts D. 2012. GSM 900 MHz radiation inhibits ants' association between food sites and encountered cues. *Electromagn. Biol. Med.* 31(2):151- 165. doi: 10.3109/15368378.2011.624661.
- Cucurachi S, Tamis WL, Vijver MG, Peijnenburg WJ, Bolte JF, De Snoo GR. 2013. A review of the ecological effects of radiofrequency electromagnetic fields (RF-EMF). *Environ. Int.* 51:116- 140.
- Di Noi A, Casini S., Campani T, Cai G, Caliani I. 2021. Review on Sublethal Effects of Environmental Contaminants in Honey Bees (*Apis mellifera*), Knowledge Gaps and Future Perspectives. *Int. J. Environ. Res. Public Health* 18:1863. <https://doi.org/10.3390/ijerph18041863>.
- Dutta SK, Subramaniam A, Ghosh B, Parshad R. 1984. Microwave radiation-induced calcium ion efflux from human neuroblastoma cells in culture. *Bioelectromagnetics NY.* 5:71- 78. <http://dx.doi.org/10.1002/bem.2250050108>.
- Edwards AW. 1998. Natural selection and the sex ratio: Fisher's sources. *Am. Nat.* 151(6):564- 569. doi:10.1086/286141.
- Fauzi A, Corebima AD. 2015. The effect of EMF radiation emitted by mobile phone to insect population using *Drosophila melanogaster* as a model organism. *Proceeding of 6th International Conference on Global Resource Conservation (ICGRC)*, Pp:16- 20.
- Fauzi A, Corebima AD, Zubaidah S. 2015. Efek Radiasi Telepon Genggam GMS terhadap Waktu Eklosi *Drosophila melanogaster*. *Proceedings of the 2nd Seminar & Workshop Nasional Biologi, IPA, dan Pembelajarannya FMIPA UM, Malang, Indonesia*, Pp:23- 26.
- Favre D. 2011. Mobile phone induced honey bee worker piping. *Apidologie.* 42:270- 279.
- Favre D, Johansson O. 2020. Does enhanced electromagnetic radiation disturb honeybees' behaviour? observations during new year's eve 2019. *Int. J. Res.-granthaalayah* 8(11):7-14. <https://doi.org/10.29121/granthaalayah.v8.i11.2020.2151>.
- Feynman RR, Sands M. 2013. *The Feynman Lectures on Physics Vol II: Mainly Electromagnetism and Matter.* Basic Books, NY.
- Galal FH, AbuElnasr A, Abdallah I, Zaki O, Seufi AM. 2017. *Culex (Culex) pipiens* mosquitoes carry and harbour pathogenic fungi during their developmental stages. *Erciyes Med. J.* 39(1):1- 6. DOI: 10.5152/etd.2017.16067.
- Gilbert LI, Rybczynski R, Warren JT. 2002. Control and Biochemical Nature of the Ecdysteroidogenic Pathway. *Annu. Rev. Entomol.* 47:883- 916.
- Goodman EM, Greenebaum B, Marron MT. 1995. Effects of electromagnetic fields on molecules and cells. *Int. Rev. Cytol.* 158:279- 338. [http://dx.doi.org/10.1016/S0074-7696\(08\)62489-4](http://dx.doi.org/10.1016/S0074-7696(08)62489-4).
- Hallmann CA, Sorg M, Jongejans E, Siepel H, Hofland N, Schwan H, Stenmans W, Muller A, Sumser H, Horren T, Goulson D, de Kroon H. 2017. More than 75 percent decline over 27 years in total flying insect biomass in protected areas. *PloS one* 12: e0185809. doi:10.1371/journal.pone.0185809.
- Hamilton WD. 1967. Extraordinary sex ratios. *Science* 156 (3774):477- 488. doi:10.1126/science.156.3774.477.
- Hardell L. 2017. Effects of Mobile Phones on Children's and Adolescents' Health: A Commentary. *Child Develop.* 89(3-4):1- 4. DOI: 10.1111/cdev.12831.
- International Telecommunication Union (ITU) 2021. *ICT Facts and Figures. ICT Data and Statistics Division, Geneva.*
- Kimmel S, Kuhn J, Harst W, Stever H. 2007. Effects of electromagnetic exposition on the behavior of the honeybee (*Apis mellifera*). *Environ. Syst. Res.* 8:1- 8.
- Kopecký K, Fernández-Martín F-D, Szotkowski R, Gómez-García G, Mikulcová K. 2021. Behaviour of Children and Adolescents and the Use of Mobile Phones in Primary Schools in the Czech Republic. *Int. J. Environ. Res. Public Health* 18:8352. <https://doi.org/10.3390/ijerph18168352>.
- Kumar NR, Sangwan S, Badotra P. 2011. Exposure to cell phone radiations produces biochemical changes in worker honey bees. *Toxicol. Int.* 18:70- 72.
- Kumar S. 2018. Colony Collapse Disorder (CCD) in Honey Bees Caused by EMF Radiation. *Bioinformation* 14(9):521- 524. doi:10.6026/97320630014521.
- Maharjan R, Yoon Y, Jang Y, Jeong M, Jung TW, Cho HS, Yi H. 2020. Artificial radiofrequency driven life-table parameters of perilla seed bugs (*Nysius* sp.) (Heteroptera: Lygaeidae). *J. Asia Pac. Entomol.* 23(4):1264- 1271.
- Maharjan R, Yi H, Ahn J, Roh GH, Park C, Yoon Y, Bae S. 2019a. Effects of radiofrequency on the development and performance of *Callosobruchus chinensis* (Coleoptera: Chrysomelidae: Bruchinae) on three different leguminous seeds. *Appl. Entomol. Zool.* 54 (3):255- 266.
- Maharjan R, Bae S, Kim GH, Yoon Y, Jang Y, Kim Y, Yi H. 2019b. Oviposition preference and development of *Marucavitrata* (Fabricius) (Lepidoptera: Crambidae) on different radiofrequency fields. *Entomol. Res.* 49 (5):214- 222. doi:10.1111/1748-5967.12349.
- Mal P, Kumar Y. 2014. Effect of electromagnetic radiation on brooding, honey production and foraging behaviour of European honey bees (*Apis mellifera* L.).

- Afr. J. Agric. Res. 9(13):1078- 1085.
- Miller GT, Spoolman SE. 2012. Living in the Environment: Principles, Connections, and Solutions. Belmont: Brooks/Cole Cengage Learning.
- Mirabolghasemi G, Azarnia M. 2002. Developmental Changes in *Drosophila melanogaster* Following Exposure to Alternating Electromagnetic Fields. *Bioelectromagnetics* 23:416-420.
- Mixson TA, Abramson CI, Nolf SL, Johnson GA, Serrano E, Wells H. 2009. Effect of GSM cellular phone radiation on the behavior of honey bees (*Apis mellifera*). *Sci. Bee Cult.* 1(2):22- 27.
- Panagopoulos DJ, Chavdoula ED, Karabarbounis A, Margaritis LH. 2007. Comparison of bioactivity between GSM 900 MHz and DCS 1800 MHz mobile telephony radiation. *Electromag. Biol. Med.* 26:33-44.
- Panagopoulos DJ, Karabarbounis A, Margaritis LH. 2004. Effect of GSM 900 MHz mobile phone radiation on the reproductive capacity of *Drosophila melanogaster*. *Electromag. Biol. Med.* 23:29- 43.
- Perrin A, Souques M. 2012. *Electromagnetic Fields, Environment and Health*. Paris: Springer-Verlag.
- Reed L. 2018. *Electromagnetic Wave Propagation*. In: *Quantum Wave Mechanics*, 3rd ed. by: Reed L., Booklocker.com, Inc., Pp: 724.
- Sainudeen SS. 2011. Electromagnetic Radiation (EMR) clashes with honey bees. *Int. J. Environ. Sci.* 1(5):897- 900.
- Samways MJ. 1993. Insects in biodiversity conservation: Some perspectives and directives. *Biodivers. Conserv.* 2(3):258- 282.
- Schowalke TD. 2006. *Insect Ecology: An Ecosystem Approach*, 2nd Ed. London: Academic Press.
- Seufi AM, Galal FH, Elsayed EH. 2007. Characterization of a *Schistocerca gregaria* cDNA encoding a novel member of mobile phone radiation-induced polypeptide related to chitinase polypeptide family. *J. App. Sci. Res.* 3(4):216- 226.
- Sharma VP, Kumar NR. 2010. Changes in honeybee behavior and biology under the influence of cellphone radiations. *Curr. Sci.* 98:1376- 1378.
- Stanojević V, Prolić Z, Savić T, Todorović D, Janać B. 2005. Effects of extremely low frequency (50 Hz) magnetic field on development dynamics of the housefly (*Musca domestica* L.). *Electromag. Biol. Med.* 24(2):99- 107.
- Sudaryadi I, Rahmawati AN, Rizqiyah M. 2020. Effect of hand phone EMF radiation on survival rate and morphological reproductive organ changes of fruit fly (*Drosophila melanogaster* Meigen, 1830). The 6th International Conference on Biological Science (ICBS 2019): "Biodiversity as a Cornerstone for Embracing Future Humanity". doi:10.1063/5.0015846.
- Taye RR, Deka MK, Rahman A, Bathari M. 2017. Effect of electromagnetic radiation of cell phone tower on foraging behaviour of Asiatic honey bee, *Apis cerana* F. (Hymenoptera: Apidae). *J. Entomol. Zool. Studies* 5(3):1527- 1529
- Thill A. 2020. Biologische Wirkungen Elektromagnetischer Felder auf Insekten. *Umwelt. Medizin. Gesellschaft.* 33:1- 28.
- Velizarov S, Raskmark P, Kwee S. 1999. The effects of radiofrequency fields on cell proliferation are non-thermal. *Bioelectrochem. Bioenerget.* 48:177- 180. [http://dx.doi.org/10.1016/S0302-4598\(98\)00238-4](http://dx.doi.org/10.1016/S0302-4598(98)00238-4).
- Wargo J, Taylor HS, Alderman N, Wargo L, Bradley JM, Addiss S. 2012. *The Cell Phones Problem: Cell Phones, Technology, Exposures, Health Effects*. North Haven: Environment and Human Health Inc.
- Weisbort D, Lin H, Ye L, Blank M, Goodman R. 2003. Effects of Mobile Phone Radiation on Reproduction and Development in *Drosophila melanogaster*. *J. Cell. Biochem.* 89:48- 55.
- World Health Organization (WHO). 2002. *Establishing a Dialogue on Risks from Electromagnetic Fields, Radiation and Environmental Health*. Geneva: Department of Protection of The Human Environment. <https://www.statista.com/statistics/262950/global-mobile-subscriptions-since-1993>.
- Xenos TD, Margas IN. 2003. Low power density RF radiation effects on experimental animal embryos and fetuses. In: Stavroulakis, P., ed. *Biological Effects of Electromagnetic Fields*. Springer. Pp:579-602.